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EFFECT OF DIFFERENT YEAST DOSES AND TIME OF APPLICATION ON GROWTH, YIELD AND QUALITY OF RUBY SEEDLESS GRAPEVINES

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ABSTRACT

This study was carried out during 2008 and 2009 seasons to investigate the effect of various yeast doses and time of application on vine surface area, yield and quality of Ruby seedless grapes. Three doses of yeast treatments (3, 6 or 9 g/vine/season) were applied via drench at three dates namely; after bud burst, after fruit set and at four weeks later.

The best effective dose was 9 g/vine/season. Application of yeast after bud burst is recommended to ensure the best vegetative growth, while, application after fruit set or four weeks later is recommended to increase the yield and enhance quality of berries i.e. enhancing, TSS/acid, total sugars content and total anthocyanins content in berry skin.

Biofertilization of Ruby seedless grapevines once with yeast at 9 g./vine/season four weeks after berry setting is essential for improving yield and quality of the berries.

M.M., Hegab *et. al.*

INTRODUCTION

The area of grape (*Vitis vinifera*, L.) increased greatly through the last decades. It reached about (١٦٧٠٤٨) feddans. The fruitful ones are about ١٥٣٩٥٦ feddans with a total annual production of (١٥٣١٤١٨) tons according to the statistics of Ministry of Agriculture and Reclamation in ٢٠٠٩. Ruby seedless grape has been commercially grown since ١٩٨٦ and extensively spread in commercial plantations, particularly in newly reclaimed areas. The berry is deep red in appearance and is very firm and sweet. So, Ruby seedless cultivar is considered as one of the most important grape for jellies, jams, wine, juice making, fresh consumption and production of raisins.

The relationship between grape yield, fruit quality and health seems to be a complex and can be influenced by nitrogen fertilization. Increasing nitrogen supply enhances photosynthesis which means that more sugar is available for growth and fruit quality (Keller, ٢٠٠٥). Whereas, excess of nitrogen supply results in an excessive vegetative growth and shaded canopies which leads to a decrease in yield and producing poor quality fruits.

Recently, a great attention has been focused on the possibility of using natural and safety substances in order to improve plant growth, yield and quality of many crops. Biofertilizers have been extensively used as an eco-friendly approach to minimize the use of chemical fertilizers, improve soil fertility status and for the enhancement of crop production by their biological activity in the rhizosphere (Ram Rao *et al.*, ٢٠٠٧).

Yeast (*Saccharomyces cervicisae*, L.) is considered as one of the promising biofertilizer for many crops (Eman *et al.*, ٢٠٠٨).

The positive effect of yeast application could be due to one or more merits: yeast aids in activating photosynthesis process through enhancing the release of carbon dioxide (Larson *et al.*, ١٩٦٢), or/and yeast contains some natural growth regulators, i.e. auxin (IAA) (Moor, ١٩٧٩) and cytokinins (Ferguson *et al.*, ١٩٨٧).

Also, the yeast was found to encourage the uptake of various nutrients as N, P and K and some common amino acids (Abou-Zaid, ١٩٨٤). Moreover, soil drench applications of yeast are probably

Biofertilization of ruby seedless grapes

promoting the uptake of different nutrient elements through modifying pH value of the soil solution towards acidity medium which reflect on yield and its components and fruit quality of various grape cultivars. In this respect, many researchers emphasized the importance of the aforementioned practices for raising yield and hence bunch quality of the vines (EL-Mogy *et al.*, 1994; Mansour, 1994; Amen *et al.*, 2006; Omran, 2006; Esmail *et al.*, 2007; Gaser *et al.*, 2007 and Abd EL-Wahab *et al.*, 2008).

Therefore, the main objective of this study was to disclose the influence of different levels from active dry yeast extract and time of application on growth, yield and fruit quality of Ruby seedless grape.

MATERIALS AND METHODS

This investigation was carried out during two successive seasons (2008 and 2009) in a private vineyard located near Gharbia governorate, on nature Ruby seedless grapevines. The vines were ten-years old, grown in a clay loamy soil under surface irrigation system, spaced at 2 x 3 meters apart and trained to the bilateral cordon. The vines were pruned during the second week of February with bud load (27 buds/vine) as 22 spurs x 2 buds. The vines were pruned to spur and trellised according to the telephone system and received the normal cultural practices usually applied in commercial orchard. Ninety uniform vines were chosen and subjected to 10 treatments with 3 replicates, 3 vines each, and each three replicates were treated with one of the following treatments :

- 1- Control.
- 2- Yeast 3 g/vine after bud burst.
- 3- Yeast 3 g/vine after fruit set.
- 4- Yeast 3 g/vine 2 weeks after fruit set.
- 5- Yeast 7 g/vine after bud burst.
- 6- Yeast 7 g/vine after fruit set.
- 7- Yeast 7 g/vine 2 weeks after fruit set.
- 8- Yeast 9 g/vine after bud burst.
- 9- Yeast 9 g/vine after fruit set.
- 10- Yeast 9 g/vine 2 weeks after fruit set.

M.M., Hegab *et al.*

All treatments were added as soil drench.

Preparation of yeast extract:

The pure dry yeast powder was activated by using sources of carbon and nitrogen with ratio of 7:1. This ratio is suitable to get the highest vegetative production of yeast, each ml of activated yeast contained about 12000 yeast cells (Barnett *et al.*, 1990). Such technique allowed yeast cells to grown multiplied efficiently during conducive aerobic and nutritional conditions. To produce de novo beneficial bioconstituents i.e. phytohormones, carbohydrates, proteins, amino acids, fatty acids, vitamins, enzymes, minerals ...etc., hence allowed such constituents to release out of yeast tissues in readily form such technique for yeast preparation based on; 1) nutritional media of glucose and casein as favourable sources of C, N and other essential elements (P, K, Mg, Fe, Mn, Cu, B and Mo as well as Na and Cl) in suitable balance (Barnett *et al.*, 1990), and 2) air pumping and adjusting incubation temperature. The media then subjected to two cycles of freezing and thawing for disruption of yeast tissues and releasing their bioconstituents directly before using. The chemical analysis of active dry yeast is shown in Table 1.

Field observations and laboratory measurements:

- 1- Average leaf area (m²/vine) : was estimated during the first week of May by picking twenty mature leaves from the fifth and seventh leaves from the shoots tip. The area was measured by using digital planimeter to measure leaf area and then multiply by leaves number/vine to determine the total leaf area/vine (m²) according to Wettstein (1901)
- 2- Chlorophyll pigments (mg/g fresh weight) : ten leaves per replicate were collected for the determination of total chlorophyll content (mg/100 g fresh weight).

Table 1: Chemical analysis of the active dry yeast according to Gaser *et al.* (2007).

N	Polysaccharides	Fats	Fiber	Ash	Thiamin (B1)	Riboflavin (B2)	Niacin (B3)	Vitamin (B6)	Vitamin (B12)
12.3%	32.3%	3.0%	1.1%	7.7%	2.33 mg	0.51 mg	37.7 mg	5.51 mg	0.02 mg

Biofertilization of ruby seedless grapes

At harvest:

- Yield (kg/vine) was estimated considering that the clusters were thinned to 4 clusters/vine.
- Cluster weight (gm).
- Total soluble solids (TSS %) was estimated using hand refractometer, total acidity (as tartaric acid in berry juice) was determined according to the A.O.A.C. (1940) and then, TSS/acid ratio was calculated. Total anthocyanin content in berry skin (mg/100 g fw) was estimated according to Hise *et al.*, (1970). Total sugar content in berries was determined by Schaffer and Somogy method as described by Ranganna (1979).

Statistical analysis :

The data obtained were statistically analyzed as complete randomized block design according to Snedecor & Cochran (1980).

RESULTS AND DISCUSSION

Vine surface area:

Data presented in Table 1 indicate that all yeast applications after bud burst improved the leaf area of Ruby seedless vines compared with the other application dates or the control. Data revealed that the highest significant values were obtained from the application of 1 g of yeast after bud burst followed by 7 and 10 g/vine in this respect amounting to 20.10-27.03-29.17, 24.43-20.19 and 22.04-24.70 m²/vine in both seasons, respectively. The positive effect of yeast applications on leaf area can be explained by the activation of photosynthesis process through enhancing the release of carbon dioxide (Larson *et al.*, 1972). Also, yeast contains natural plant growth promoters specially IAA and cytokinins (Moor, 1979).

The present results are in line with those obtained by Mansour (1994) on Anna apple, El-Mogy *et al.*, (1994) on Thompson seedless cv., Amen *et al.*, (2000a) on King Ruby cv. Omran (2000) and Esmaeil *et al.*, (2003) on Roumi Red cv. and Gaser *et al.*, (2007) on Flame seedless grape who pointed out that yeast application as foliar or soil

drench significantly increased vegetative growth compared with the untreated vines (control).

Table 7: Effect of yeast doses and time of application on leaf area and yield during 2007 and 2008 seasons.

Treatment	Leaf area (m ² /vine)		Yield (kg/vine)	
	2007	2008	2007	2008
Control	18.45	19.21	18.0	18.8
Yeast 7 g/vine after bud burst.	22.54	24.75	19.1	19.6
Yeast 7 g/vine after fruit set.	19.40	20.40	20.0	20.0
Yeast 7 g/vine 4 weeks after fruit set.	19.37	20.40	21.0	21.0
Yeast 7 g/vine after bud burst.	24.43	25.91	20.6	20.8
Yeast 7 g/vine after fruit set.	19.64	20.86	21.0	21.3
Yeast 7 g/vine 4 weeks after fruit set.	19.59	20.84	22.8	22.8
Yeast 9 g/vine after bud burst.	26.53	29.16	21.8	21.9
Yeast 9 g/vine after fruit set	20.13	20.93	22.3	22.3
Yeast 9 g/vine 4 weeks after fruit set.	20.15	21.02	23.2	23.2
L.S.D at 5 %	0.41	0.53	0.28	0.31

Yield :

Data in Table 7 show that all yeast treatments significantly increased the yield of King Ruby cv. compared with the control. Yeast applied at 9 g/vine 4 weeks after fruit set resulted in the highest values (23.2-23.2 kg/vine) followed by 7 g/vine (22.8-22.8 kg/vine) during both seasons, respectively. This positive effect can be explained by the activation of photosynthesis process through enhancing carbon dioxide release, Furthermore, and IAA and cytokinin- like substance which encourage the uptake of various nutrients (Moor, 1979).

These results are nearly similar to those reported by Amen *et al.*, (2007a) on King Ruby and Gaser *et al.*, (2007) on Flame seedless who found that yeast applications significantly increased the yield/vine.

TSS, acidity and TSS/acid ratio:

Data in Table 7 indicated that yeast application 4 weeks after fruit set resulted in more pronounced values of TSS %, TSS/acid ratio

Biofertilization of ruby seedless grapes

and decreased acidity compared with the control. Applied doses significantly increased the parameters compared with the control except for the acidity which significantly decreased.

Table 7: Effect of yeast doses and time of application on TSS, acidity and TSS/acid ratio during 2008 and 2009 seasons.

Treatment	TSS %		Acidity %		TSS/acid ratio	
	2008	2009	2008	2009	2008	2009
Control	10.0 3	10.7	0.36	0.30	42.7	44.8
Yeast 2 g/vine after bud burst	16.2 .	16.0	0.22	0.21	72.0	78.4
Yeast 2 g/vine after fruit set	16.6 7	16.9	0.22	0.20	74.6	83.0
Yeast 2 g/vine 4 weeks after fruit set.	10.7 .	18.2	0.24	0.23	72.3	76.4
Yeast 7 g/vine after bud burst	16.4 .	16.8	0.30	0.29	04.1	07.9
Yeast 7 g/vine after fruit set	16.8 7	17.0	0.31	0.30	04.4	09.0
Yeast 7 g/vine 4 weeks after fruit set	17.7 3	17.9	0.27	0.20	66.6	72.7
Yeast 9 g/vine after bud burst	17.0 .	17.0	0.29	0.26	08.0	64.6
Yeast 9 g/vine after fruit set	17.8 .	18.6	0.28	0.27	64.4	69.7
Yeast 9 g/vine 4 weeks after fruit set	18.8 .	19.3	0.28	0.27	67.9	71.3
L.S.D at 0 %	0.06	0.48	0.02	0.01	0.73	4.48

Table 7 also reveals that the highest significant effect was in the application of 9 g yeast 4 weeks after fruit set amounting to 18.8-19.3

M.M., Hegab *et. al.*

% in the case of TSS %. Meanwhile, in the case of acidity % and TSS/acid ratio the results did not take the same line but differed from one to another. The positive effects of yeast application on berry chemical properties i.e. TSS %, TSS/acid ration and the negative effects on acidity % in the grape juice could be attributed to the enhancement effects of photosynthesis processes and increasing promoters hormones as cytokinins (Moor, 1979).

It is well known that these hormones induce a considerable amount of sugar contents and consequently caused an increase in TSS %, TSS/acid ratio and a decrease in acidity % in the grape juice. These results are in agreement with those found by Gaser *et al.*, (2007) who found that yeast application increased TSS, TSS/acid ratio and decreased total acidity of the juice of Flame seedless.

Total sugars, anthocyanin and chlorophyll:

Data in Table 4 show that all treatments increased total sugars, total anthocyanin and chlorophyll compared with the control. Yeast application (7 g/vine) 4 weeks after fruit set resulted in the highest significant sugars content, anthocyanin and chlorophyll than the other treatments or the control.

Table 4: Effect of yeast doses and time of application on total sugars, total anthocyanin and chlorophyll during 2008 and 2009 seasons.

Treatment	Total sugars %		Total anthocyanin (mg/100g)		Chlorophyll (mg/100g fw)	
	2008	2009	2008	2009	2008	2009
Control	12.60	12.82	2.14	2.19	18.40	19.21
Yeast 7 g/vine after bud burst	12.97	13.14	2.69	2.71	22.04	24.60
Yeast 7 g/vine after fruit set	13.38	13.87	2.72	2.73	19.40	20.40
Yeast 7 g/vine 4 weeks after fruit set.	13.97	13.99	2.74	2.76	19.37	20.40
Yeast 7 g/vine after bud burst	13.91	14.00	2.71	2.78	24.43	20.91
Yeast 7 g/vine after fruit set	14.64	14.89	3.01	2.92	19.64	20.86

Biofertilization of ruby seedless grapes

Yeast 7 g/vine 4 weeks after fruit set	15.83	15.13	3.11	3.13	19.59	20.84
Yeast 4 g/vine after bud burst	14.45	14.15	2.93	2.83	26.53	29.16
Yeast 4 g/vine after fruit set	14.21	15.81	3.16	3.17	20.13	20.93
Yeast 4 g/vine 4 weeks after fruit set	17.39	17.30	3.21	3.30	20.15	21.02
L.S.D at 5 %	0.23	0.36	0.07	0.19	0.41	0.53

M.M., Hegab et. al.

The highest values were obtained from the application of 9 g yeast 4 weeks after fruit set amounting to 14.37-14.30 % in the case of total sugars, 3.21-3.30 mg/g in the case of total anthocyanin and the highest values of chlorophyll were obtained from the application of 9 g yeast after bud burst 27.03-29.17 mg/g fw in the both seasons, respectively. These results are in agreement with those found by Amen et al., (2008a) on King Ruby, Gaser et al., (2007) on Flame seedless and Abd EL-Wahab et al., (2008) on Black Monukka grapevine.

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M.M., Hegab *et. al.*

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Biofertilization of ruby seedless grapes

تأثير تركيز ومواعيد إضافة الخميرة الجافة على النمو والمحصول وبعض صفات جودة الثمار لكرمات عنب الكنج روبي

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أجرى هذا البحث بغرض دراسة تأثير المعاملة بالخميرة الجافة بتركيزات مختلفة ومواعيد مختلفة وذلك على المسطح الورقي للكرمة (متر مربع) وكمية المحصول وجودة حبات العنب الروبى سيدلس. تم إضافة ثلاثة جرعات من الخميرة الجافة أرضياً وهى (٣ ، ٦ ، ٩ جم/كرمة) فى ثلاثة مواعيد مختلفة وهى : الأول بعد إكمال تفتح البراعم ، والثانى بعد العقد مباشرة ، والثالث بعد العقد بأربعة أسابيع.

وقد أوضحت النتائج المتحصل عليها من هذه الدراسة إلى أن إضافة الخميرة بتركيز ٩ جم/كرمة أعطى أفضل النتائج حيث أنه عند إضافتها فى الموعد الأول أعطت أفضل مسطح ورقي أما عند إضافتها فى الموعدين الثانى والثالث أعطت أعلى محصول/كرمة بالإضافة إلى تحسين خصائص الحبات فى صورة زيادة المواد الصلبة الذائبة الكلية ونسبتها إلى الحموضة والسكريات الكلية فى الحبات وصبغة الأنثوسيانين فى قشرة الحبات.

ان التسميد الحيوي لكرمات العنب الروبى سيدلس باستخدام الخميرة بمعدل ٩ جرام/كرمة/ موسم مرة واحدة بعد عقد الحبات بأربعة أسابيع يكون ضروريا لتحسين كمية المحصول وجودة الحبات.